Human 62.5pg/ml – 4000 pg/ml No detectable cross-reactivity with other cytokines

KIT COMPONENTS

Component	Amount
96-well plate precoated with anti-human Survivin antibody	1 Plate
Protein Standard: Lyophilized recombinant human Survivin	2 tubes, 4 ng/tube
Sample Diluent Buffer	30 ml
Biotinylated Antibody (Anti-human Survivin)	130 µl (100x)
Antibody Diluent Buffer	12ml
Avidin-Biotin-Peroxidase Complex (ABC) Solution	130 µl (100x)
ABC Diluent Buffer	12 ml
Tetramethyllbenzidine (TMB) Color Developing Agent	10 ml
TMB Stop Solution	10 ml

Storage

Store at 4°C. Cell Applications, Inc. recommends using the kit within 6 months of order.



graphed against Survivin protein concentration. The TMB reaction was incubated at 37° C for 22 min.

BACKGROUND

Survivin, a novel human number of the inhibitor of apoptosis protein (IAP) family, has been suggested to directly inhibit caspase-3 and -7 activity or conjugate caspase-9, and regulate the G2/M phase by interact with spindle microtubules. Survivin is expressed in fetal development, in normal endometrium primarily during the secretory phase of the menstrual cycle and in cancers, whereas no transcripts were detected in terminally differentiated adult tissues. Expression of survivin on breast, neuroblastoma, lung, esophageal, and colorectal cancers correlates with an unfavorable prognosis, which is shortened survival and/or a shortened time to recurrence.1 Besides its role as an IAP, survivin acts as a subunit of the chromosomal passenger complex (CPC) and as a regulator of microtubule dynamics. The typical chromosomal passenger localization pattern of survivin can be observed not only in normal but also in tumor cells.2 In normal cells, expression of survivin peaks at the G2/M transition of the cell cycle, whereas the cell cycle-dependent transcriptional control of survivin is deregulated by various oncogenic pathways in cancer cells. Hence, survivin is found in the majority of interphase tumor cells in patients, which supports its bifunctional role. Moreover, the observation that survivin can be found not only in the cytoplasm but also in the nucleus of tumor and proliferating normal cells (e.g., endothelial and CD34+ stem cells) stimulated the hypothesis that these subcellular pools may coincide with different survivin functions. Nuclear survivin was suspected to control cell division, whereas cytoplasmic/mitochondrial survivin was considered cytoprotective. A failure in passing on the duplicated genetic material to both daughter cells together with resistance to apoptosis can contribute to cellular transformation and cancer progression. Survivin seems to intersect both processes.3

References

1. Altieri, D.C.: Trends Mol. Med. 7:542-7, 2001 2. Xia, F. et al: Mol. Cell. Biol. 28:5299-5311, 2008 3. Stauber, R.H. et al: Cancer Res. 67:5999-6002, 2007

ELISA OVERVIEW

Cell Applications ELISA Kits are based on standard sandwich enzymelinked immunosorbent assay technology. Freshly prepared standards, samples, and solutions are recommended for best results.

- 1. Prepare test samples.
- 2. Prepare a protein standard of the target protein.
- 3. Add test samples and standards to the pre-coated 96-well plate. Do not wash.
- 4. Add biotinylated detection antibodies. Wash.
- 5. Add Avidin-Biotin-Peroxidase Complex (ABC) Solution. Wash.
- 6. Add Tetramethyllbenzidine (TMB) Color Developing Agent,
- containing HRP substrate. 7. Add TMB Stop Solution
- 8. Subject the plate to analysis.

NOTES:

- Before using the kit, quick spin tubes to bring down all solution to the bottom of tube.
- Duplicate assay wells are recommended for both standard and sample testing.
- Do not let the 96-well plate dry, this will lead to inactivation of plate components.
- When diluting samples and reagents, ensure that they are mixed completely and evenly.
- Pre-warm diluted ABC and TMB solutions at 37°C for 30 min before use to avoid variable temperature effects.
- For washes, use TBS or PBS. Do not touch well walls.
- A protein standard is included in the kit. A protein standard detection curve should be generated with each experiment, no more than 2 hours prior to the experiment.
- The user will determine sample dilution fold by estimation of target protein amount in samples.

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ELISA PROTOCOL

Preparation of Test Samples

1. Process Test Samples in the following manner:

- Cell culture supernate, tissue lysate or body fluids: Remove particulates by centrifugation, analyze immediately or aliquot and store at -20°C.
- Serum: Allow the serum to clot in a serum separator tube (about 3 0 m i n) at room temperature. Centrifuge at approximately 1500 x g for 15 min. Analyze the serum immediately or aliquot and store frozen at -20° C.
- **Plasma**: Collect plasma using heparin or EDTA as an anticoagulant. Centrifuge for 15 min at 1500 x g within 30 min of collection. Analyze immediately or aliquot and store samples at -20°C.
- 2. Estimate the concentration of the target protein in the sample and select a proper dilution factor such that the diluted target protein concentration falls within the 62.5-4000 pg/ml standard curve range. Depending on the sample, several trial dilutions may be necessary. Dilute the sample using the provided diluent buffer, mixing well. Suggested working dilutions of samples are as follows:

Target Protein	Sample	Sample	Diluent
Concentration Range	Working Dilution	Vol.	Buffer Vol.
40-400 ng/ml	1:100	1 µl	99 µl
4-40 ng/ml	1:10	10 µl	90 µl
62.5-4000 pg/ml	1:2	50 µl	50 µl
<62.5 pg/ml	n/a	100ul	n/a

 If samples will be assayed within 24 hours, store at 2-8° C. For long-term storage, aliquot and freeze samples at -20° C. Avoid repeated freeze-thaw cycles.

Preparation of Standard Solutions (62.5-4000 pg/ml)

- 4. Add 1 ml of sample diluent buffer into one tube, keep the tube at room temperature for 10 min and mix to make a 4000 pg/ml Survivin solution.
- Label 6 eppendorf tubes with the Survivin protein concentrations to be prepared by serial dilution: 2000pg/ml, 1000pg/ml, 500pg/ml, 250pg/ml, 125pg/ml, 62.5pg/ml.
- 6. Aliquot 0.3 ml of the Sample Diluent Buffer to the labeled tubes.
- 7. Serially dilute the protein standards into their respectively labeled tubes. Transfer 0.3 ml from the 4000pg/ml Survivin Solution to the 2000pg/ml eppendorf tube and mix thoroughly. Transfer 0.3 ml of the 2000 pg/ml solution to the 1000pg/ml tube and mix thoroughly, Transfer 0.3 ml of the 1000pg/ml solution to the 500pg/ml tube and mix, and so on to make the 250, 125, and 62.5 pg/ml solutions.
- 8. Store at 4° C until use.



Loading the 96-well Plate

- 9. Aliquot 0.1 ml of the sample diluent buffer into a control well to serve as the Blank. This will yield the O.D.450(Blank) reading.
- Aliquot 0.1 ml of the standard solutions of the Preparation of Standard Solutions (62.5-4000 pg/ml) into empty wells of the precoated 96-well plate. Duplicate measurements of standards are recommended.

- 11. Aliquot 0.1 ml of each properly diluted test sample to empty wells prepared in **Step 2**. Duplicate measurements of each test sample are recommended.
- 12. Cover the 96-well plate and incubate at 37° C for 90 min.
- 13. During the Step 12 incubation period, prepare a stock of Biotinylated 1:100 Antibody Working Solution. Count the number of reactions and multiply by 0.1 ml/well for the Working Solution total volume (preparation of 1-2 reactions in excess of the number of wells is recommended). Dilute the Biotinylated Antibody to 1:100 in Antibody Diluent Buffer and mix thoroughly. Use the working solution within 2 hours.
- 14. Upon completion of the 90 min incubation of **Step 12**, remove the cover of the 96 well plate and discard plate well contents. Blot the plate onto paper towels or other absorbent material. DO NOT let the wells completely dry at any time.
- Add 0.1 ml of the Biotinylated 1:100 Antibody Working Solution (prepared in Step 13) to each well and incubate the plate at 37° C for 60 min.
- 16. During the incubation period of Step 15, prepare a stock of ABC Working Solution. Count the number of reactions and multiply by 0.1 ml/well for the Working Solution total volume (preparation of 1-2 reactions in excess of the number of wells is recommended). Dilute the ABC Stock Solution to 1:100 in ABC Diluent Buffer and mix thoroughly. Pre-warm the ABC working solution at 37° C for 30 min before use. Use the working solution within 1 hour.
- 17. Upon completion of the 60 min incubation of **Step 15**, wash the plate 3 times with 0.3 ml TBS or PBS. For each wash, leave washing buffer in the wells for 1-2 min. Discard the washing buffer and blot the plate onto paper towels or other absorbent material.
- Add 0.1 ml of prepared ABC Working Solution (prepared in Step 16) to each well and incubate the plate at 37° C for 30 min.
- During the incubation period of Step 18, pre-warm TMB Color Developing Agent at 37° C for 30 min before use.
- 20. Upon completion of the 30 min incubation of Step 18, wash the plate 5 times with 0.3 ml TBS or PBS. For each wash, leave the washing buffer in the wells for 1-2 min. Discard the washing buffer and blot the plate onto paper towels or other absorbent material.
- Add 90 µl of the pre-warmed TMB Color Developing Agent into each well and incubate at 37° C for 20-25 min (shades of blue can be seen in the wells with the four most concentrated Protein Standard Solutions; the other control wells should show no obvious color).
- 22. Add 0.1 ml of the TMB Stop Solution to each well. The acidic stop solution will change the mixture color to yellow. The yellow intensity is proportional to the amount of target protein captured by the plate.
- Read the O.D. absorbance at 450nm in a microplate reader within 30 min after adding the stop solution. These readings are the O.D.450(Reading).

Calculating Protein Concentration

- For all wells, determine O.D.450(Relative): O.D.450(Relative) = O.D.450(Reading) – O.D.450(Blank)
- Plot the standard curve: plot O.D.450(Relative) of each standard solution (Y) vs. the respective concentration of the standard solution (X). See Figure 1 for a typical standard curve.
- The Survivin concentration of the samples can be interpolated from the standard curve. Multiply the interpolated concentration by the dilution factor to obtain the target protein concentration in the sample.

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